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THE CHEMICAL PROPERTIES OF N-(O,O-DIISOPROPYL)PHOSPHORYL-ARGININE

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N-(O,O-diisopropyl)phosphoryl-arginine (DIPP—Arg) **3** and N-(O,O-diisopropyl)phosphoryl-arginine methyl ester (DIPP—Arg—OMe) **4a** were synthesized. In compound **3** the co-participation of the three functional groups of the phosphoryl, guanidine and carboxyl was essential for the ester exchange reaction at the phosphoryl group and esterification at carboxyl group. In contrast, when the carboxyl group was blocked, as in compound **4a**, no corresponding reaction occurred. A mixed anhydride penta-coordinate phosphorus transition state was proposed for the self-activation mechanism.

Key words: Phosphorylated amino acid, N-phosphorylated arginine, ester exchange on phosphate.

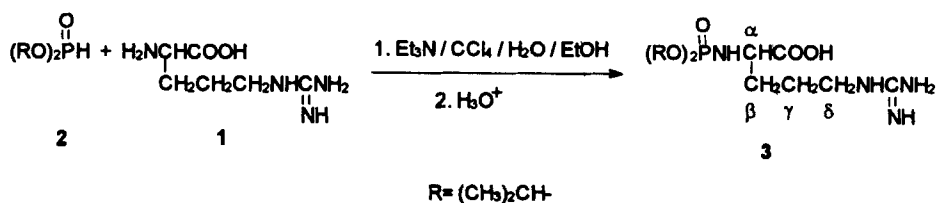
INTRODUCTION

A novel synthesis of N-phosphorylated amino acids was found by Zhao *et al.*,¹ and peptide formation, esterification and ester exchange reactions of these compounds were studied.^{2–3} In terms of basic amino acids, such as arginine, histidine and lysine, there were some problems in obtaining the pure phosphoryl products. Therefore, compound **3** was synthesized through a modified method, and its properties were studied.

RESULTS AND DISCUSSIONS

Since there were two basic functional groups such as amino and guanidine groups present in arginine **1**, if diisopropylphosphite (DIPPH) **2** was in excess, both amino groups were phosphorylated. The ratio of DIPPH **2** and arginine **1** was maintained at 1:1, and the reaction temperature was kept below -10°C for at least 30 minutes. Only the α -amino group was phosphorylated (Scheme 1).

Compounds **4a** and DIPP—Ala¹ **9** were also synthesized. The purity and structure of compounds **3** and **4a** were determined by fast atom bombardment mass spectrometry (FAB-MS), HRFAB-MS, $^1\text{H-NMR}$, $^{31}\text{P-NMR}$ and $^{13}\text{C-NMR}$.



SCHEME 1. Synthesis of DIPP—Arg 3

Ester Exchange and Esterification Reactions

A solution of compound 3 in 1-butanol was kept at 40°C for 20 hr. The FAB-MS of the mixture indicated that in addition to the molecular peak at $(M + 1)^+ / z$ 339 for compound 3, there were several new signals at $(M + 1)^+ / z$ 395, 409, 423, 353, 367, which corresponded to the products from mono- and di-ester exchange at the phosphoryl group, i.e., N—(iPrO)(BuO)P(O)—Arg 5b and N—(BuO)₂P(O)—Arg 6b, together with the esterification products DIPP—Arg—OBu 4b, N—(iPrO)(BuO)P(O)—Arg—OBu 7b, N—(BuO)₂P(O)—Arg—OBu 8b, respectively (Table I). These compounds showed overlapping ³¹P-NMR signals at 6.27, 7.36, 8.46 ppm. According to the calculated ³¹P-NMR shifts for N-phosphoryl amino acids,⁷ the ³¹P-NMR shifts for compounds 3, 5b and 6b should be 6.27, 7.27 and 8.27 ppm, respectively. Since the ³¹P-NMR shifts of N-phosphoryl amino acid and its ester were almost the same, they gave an overlapping signal. The relative amount of each type of compound were reported as the title ³¹P-NMR peak intensity.

Similarly, after keeping freshly prepared compound 3 in methanol at 40°C for 20 hours, the compounds listed in Table II also showed overlapping ³¹P-NMR shift values at 6.45, 8.80, 11.18 ppm, respectively. These deviated somewhat from the calculated values of 6.45, 8.25, 10.05 ppm. All these reactions are shown in Scheme 2.

Mechanistic Study

When DIPP—Arg—OMe 4a was dissolved in 1-butanol or methanol and kept at 40°C for 20 hr, the ³¹P-NMR spectrum indicated that no similar reaction occurred.

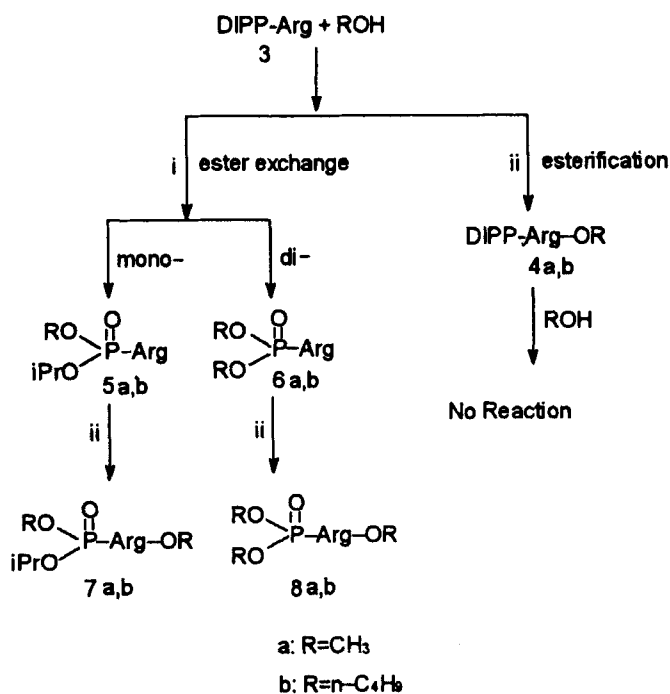
TABLE I
The FAB-MS and ³¹P-NMR data of DIPP—Arg 3 after reacted with 1-butanol at 40°C for 20 hr

(M+1) ⁺ /z	³¹ P-NMR (ppm) & relative amount	possible compd.	ester exchange	esterification
339 395=339+56	6.27 (28.2%)	DIPPArg DIPPArgOBu		ester
353=339+14 409=353+56	7.36 (44.0%)	(BuO)(iPrO)P(O)Arg (BuO)(iPrO)P(O)ArgOBu	mono- mono-	ester
367=339+28 423=367+56	8.46 (27.8%)	(BuO) ₂ P(O)Arg (BuO) ₂ P(O)ArgOBu	di- di-	ester

TABLE II

The FAB-MS and ^{31}P -NMR data of DIPP—Arg 3 after reacted with methanol at 40°C for 20 hr

$(M+1)^+/z$	^{31}P -NMR (ppm) & relative amount	possible compd.	ester exchange	esterification
339 353=339+14	6.45 (38.9%)	DIPPArg DIPPArgOCH ₃		ester
311=339-28 325=311+14	8.80 (40.8%)	(CH ₃ O)(iPrO)P(O)Arg (CH ₃ O)(iPrO)P(O)ArgOCH ₃	mono- mono-	ester
283=339-56 297=283+14	11.18 (20.3%)	(CH ₃ O) ₂ P(O)Arg (CH ₃ O) ₂ P(O)ArgOCH ₃	di- di-	ester

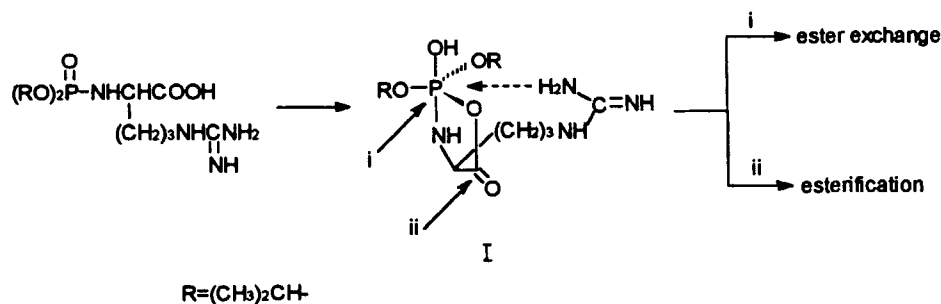


group, because a simple phosphoryl amino acid without a side chain functional group, such as DIPP—Ala **9** had much lower reactivity. Furthermore, on addition of guanidine to the solution of DIPP—Ala **9** in methanol, there was no intermolecular base catalysis. On the contrary, the ester exchange and esterification reactions were completely inhibited. This implies that the guanidine group was not a general base catalysis, but an intramolecular catalytic group.

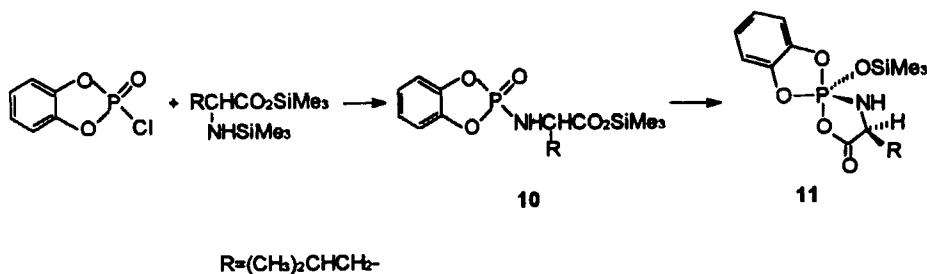
To account for these facts, the co-participation of all three groups in the DIPP—Arg **3** is proposed for the esterification and ester exchange reactions (Scheme 3). The transition state was a pentacoordinate phosphorus compound, and it could be subsequently activated by the intramolecular guanidine group.

In this reaction, the lifetime of the transition state was too short to be trapped by ^{31}P -NMR spectroscopy. In the following reaction, a model compound, leucine derivative **10**, with a stabilizing trimethylsilyl group $(\text{Me})_3\text{SiO}-$ was found to be transferred into the pentacoordinate product **11** completely (Scheme 4). Compound **11** was stable and its structure was identified. But for DIPP—Arg **3**, the derivative corresponding to compound **11** could not be trapped by ^{31}P -NMR yet. However, the identification of compound **11** might give some support to the proposed mechanism.

DIPP—Leu **13** could also undergo the same reactions as DIPP—Arg **3** at a much lower reactivity, because there was no side chain catalytic group in compound **13**.



SCHEME 3. Proposed transition state for the reaction of DIPP—Arg **3** with alcohol



SCHEME 4. Synthesis of transition state (I) analogue—compound **11**

CONCLUSION

As mentioned above, since the phosphoryl group in N-DIPP-Arg-OMe **4b** was inert, neither the esterification nor ester exchange occurred. However, in DIPP-Arg **3** with a free carboxyl to activate the phosphoryl group, together with an intramolecular guanidine catalytic effect, the phosphoryl ester exchange and esterification reactions were possible. Esterification and ester exchange at the phosphoryl group are fundamental biochemical reactions. Esterification of the amino acid is the first step in protein bio-synthesis. Ester exchange at the phosphoryl group is the key of the self-catalysis of RNA, and this has been postulated to relate to the foundation of biocatalysis.⁴⁻⁶ In many cases enzymes are activated by a phosphorylation mechanism, and the position of phosphorylation in protein was found to be mostly in the arginine, histidine, serine, threonine and tyrosine residues. The results of this paper might provide a clue to arginine's side chain catalytic function in enzymes.

EXPERIMENTAL

Methods

The ¹H-NMR, ¹³C-NMR and ³¹P-NMR spectra were taken on a JEOL-JNM FX-100 and BRUKER AC200P spectrometer. The positive-ion FAB-MS data were obtained on a KYKY ZHP-5 double-focusing mass spectrometer, equipped with a standard KYKY fast atom gun. Infrared spectra were determined with a Carlzeiss, Jena Sepeord 751R instrument. EI and HR EI-MS data were taken on a ZAR-HS GC-MS spectrometer.

Preparation of N-(diisopropylphosphoryl)-arginine **3** (DIPP-Arg)

To an ice-salt cooled solution of arginine mono-hydrochloride (2.107 g, 10 mmol) in Et₃N (7 ml), H₂O (10 ml) and EtOH (5 ml) a mixture of diisopropyl phosphite **2** (1.66 g, 10 mmol) and CCl₄ (5 ml) was added dropwise. The mixture was stirred at -15°C for 2 hr and R.T. for 30 min. The mixture was evaporated in vacuum and the residue washed with ether (2 × 20 ml). The water layer was adjusted to pH 3-4 by 1.5 N HCl and extracted with 1:2 tertbutyl alcohol-ethyl acetic ester (5 × 20 ml). The combined extracts were washed with saturated aqueous NaCl, dried (MgSO₄) and concentrated in vacuum. A colorless solid product **3** was obtained, yield 72%. ³¹P-NMR: 6.11 ppm (BuOH-CH₃COOEt). ¹H-NMR: (d₆-DMSO) 1.28 (d, 12H, *J* = 6 Hz, 4 × CH₃), 1.60 (m, 4H, β, γ-CH₂), 3.17 (m, 2H, δ-CH₂), 3.55 (m, 1H, NH-P), 4.52 (m, 3H, CH-O and α-CH), 7.28 (m, 3H, H₂N-C=NH), 7.92 (m, 1H, -C-NH), 11.5 (s, 1H, -COOH). ¹³C-NMR (d-DMSO): 23.6 (-CH₃), 37.8 (β-CH₂), 31.3 (γ-CH₂), 55.0 (δ-CH₂), 67.4 (α-CH), 69.8 (-CH-O), 157.5 (-NH-C=NH), 177.0 (-COOH). Positive FAB-MS: (M + 1)⁺/*z* = 339. IR: 1652, 1438, 1388, 1000, 3400, 3320, 3240, cm⁻¹. HRFAB-MS: molecular formula C₁₂H₂₇N₄PO₅, (M + 1)⁺/*z* = 339.1797 (calculated), 339.1794 (measurement).

Preparation of N-DIPP-Arg-OMe **4b**

To an ice-salt cooled solution of Arg-OMe·2HCl (1.3 g, 5 mmol) in methanol (10 ml) and Et₃N (2.8 ml) a mixture of diisopropyl phosphite **2** (1 ml, 6 mmol) and CCl₄ (10 ml) was added dropwise. The mixture was stirred for 10 hr, then evaporated. The residue was dissolved in saturated aqueous NaCl and washed with petroleum ether (3 × 20 ml). Then the water layer was extracted with ethyl acetate ester (3 × 20 ml), and the combined extracts were washed with saturated aqueous NaCl and dried (MgSO₄) and concentrated in vacuum. A colorless oil **4b** was obtained, yield 85%. ³¹P-NMR: 5.36 ppm. ¹H-NMR: (d₆-DMSO) 1.32 (d, 12H, *J* = 6 Hz, 4 × CH₃), 1.77 (m, 4H, β, γ-CH₂), 3.32 (m, 2H, δ-CH₂), 3.75 (s, 3H, -OCH₃), 3.8 (m, 2H, α-CH and NH-P), 4.54 (m, 2H, CH-O), 7.28 (m, 3H, H₂N-C=NH), 7.92 (m, 1H, -C-NH). ¹³C-NMR (d-DMSO): 23.6 (-CH₃), 40.7 (β-CH₂), 33.5 (γ-CH₂), 52.3 (δ-CH₂), 53.5 (α-CH), 71.6 (-CH-O), 157.2 (-NH-C=NH), 173.3 (-COOH), 77.6 (-OCH₃). IR (film): 3156, 1740, 1665, 1220, 990 cm⁻¹. FAB-MS: (M + 1)⁺/*z* = 353. HRFAB-MS: for molecular formula C₁₃H₂₉N₄PO₅, (M + 1)⁺/*z* = 353.2001 (measurement), 353.1983 (calculated).

Preparation of compound 11

All operation was under protection of nitrogen. 0.4 ml 0.5 M N,O,O-tri(methylsilyl) leucine⁸ in CCl₄ and 0.2 ml 1M O,O-phenylene phosphochloridate⁹ in CCl₄ were injected into a 5 mm NMR tube. The tube was sealed immediately and the reaction was traced by ³¹P-NMR. First, the phosphoryl amino acid **10** was formed, (³¹P-NMR: 21.3 ppm) and then it was transferred into the penta-coordinate product **11** completely as checked by ³¹P-NMR. The solvent was removed in vacuum and the colorless oil product **11** was obtained, yield almost 99%. ³¹P-NMR: -45.1, -45.6. ¹H-NMR: 0.43 (s, 9H), 0.91-1.02 (q, 6H), 1.71-1.82 (m, 1H), 3.83-3.91 (m, 1H), 4.56 (s, 1H), 6.75-7.04 (m, 4H). ¹³C-NMR: 0.82, 21.71, 23.01, 25.18, 42.86, 53.35, 100.86, 110.38, 111.16, 111.95, 120.27, 123.30, 169.50. EI(m/z): 357 (C₁₅H₂₄NO₅PSi). HR EI-MS: 357.1165 (measurement), 357.116 (calculated).

Preparation of DIPP-Ala **9**, DIPP-NH-Bu **12**, DIPP-Leu **13** is referred to reference 1.

Acetic acid action on DIPP-NH-Bu **12**

60 mg (1 mmol) acetic acid was added into 1 ml 0.5 M compound **12** solution in methanol, then the solution was kept at 40°C for 20 hr. ³¹P-NMR spectra showed that there was no reaction occurred.

Guanidine action on DIPP-Ala **9**

60 mg(1 mmol) guanidine was added into 1 ml 0.5 M compound **9** solution in methanol, then the solution was kept at 40°C for 20 hr. ³¹P-NMR spectra showed that the ester exchange and esterification reactions were completely inhibited.

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